

In-vitro/ex-vivo exposure system dosimetry: successes and challenges

Symposium: In Vitro Test Methods
to Model Local Respiratory Effects
after Exposure to Pulmonary
Toxicants: Not just
smoke and mirrors
SOT Annual Meeting
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Michael J. Oldham

Conflict of Interest Disclosure

- Employee of Altria Client Services LLC
- Altria Client Services LLC has purchased in vitro ALI equipment from Vitrocell Systems GmbH



Outline

- Goal
- Submerged Cultures
- Air-Liquid Interface – Successes
- Air-Liquid Interface – Challenges
- Air-Liquid Interface - Opportunities

Goal

- Know the internal cell dose that results in the observed in-vitro/ex-vivo response?
- Know the cell surface dose that results in the observed in-vitro/ex-vivo response?
- Know the cell exposure concentration that results in the in-vitro/ex-vivo observed response?
- Know the intended exposure concentration that results in the in-vitro/ex-vivo observed response?



Submerged Cultures - Successes

- ISDD: (In-vitro Sedimentation, Diffusion Dosimetry) model by Hinderliter et al., 2010 & ISD³ Thomas et al., 2018

R = gas constant

T = temperature

Na = Avogadro's #

μ = media viscosity

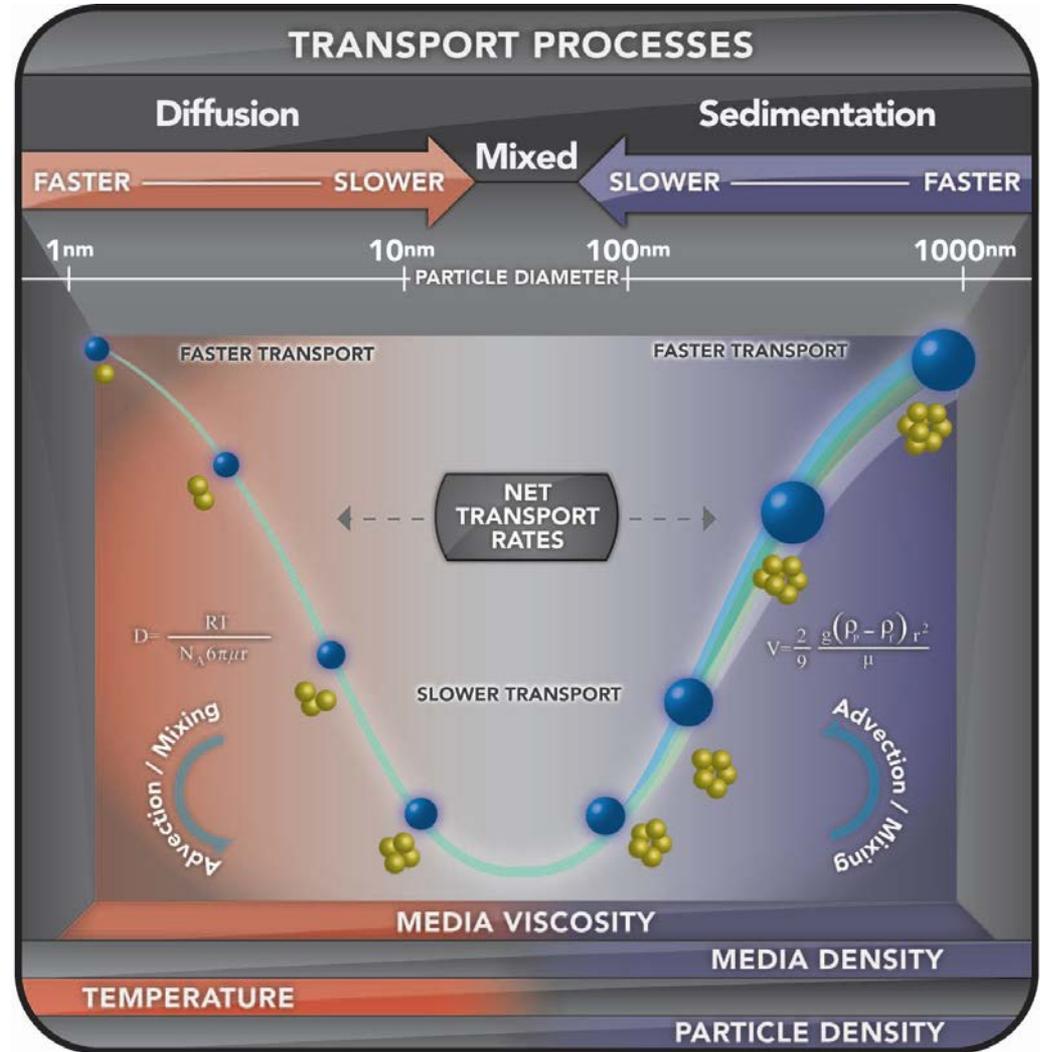
r = particle radius

g = gravitational acceleration

ρ_p = particle density

ρ_f = fluid density

L = total media height

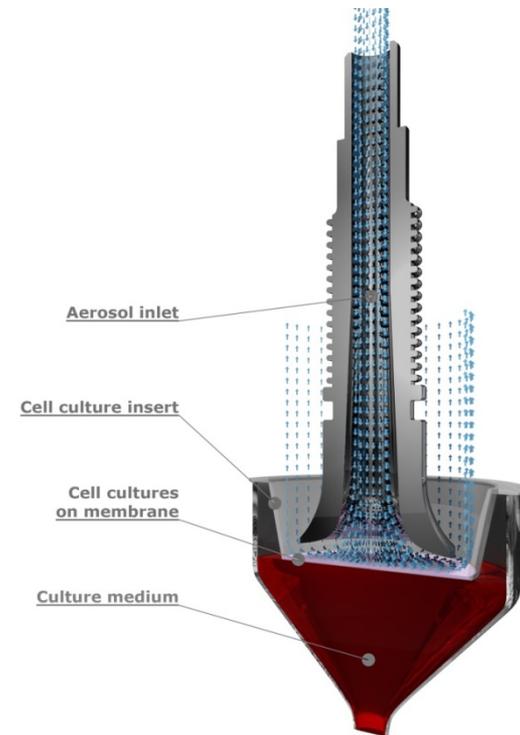
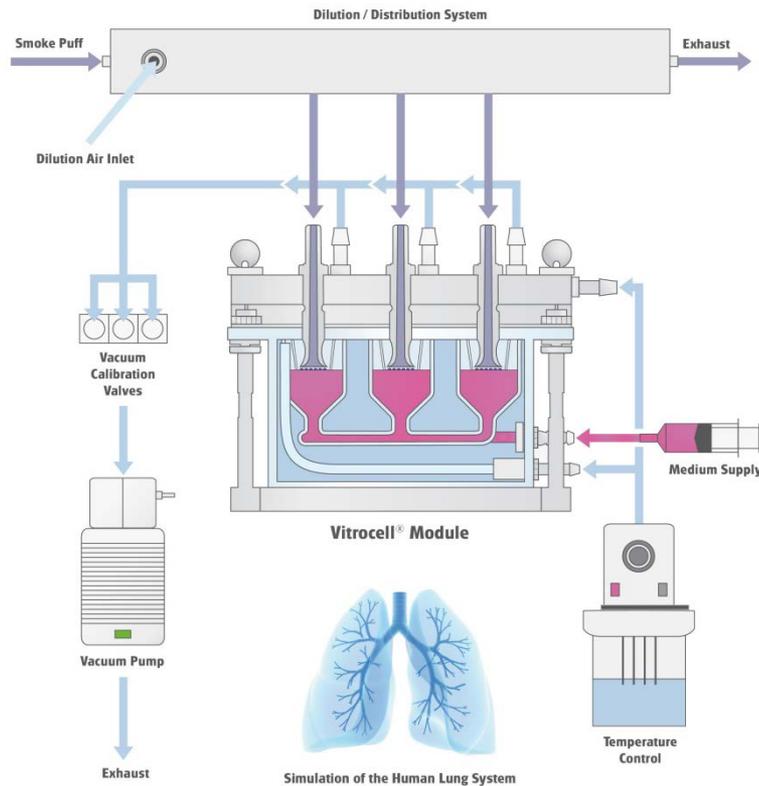


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Air-Liquid-Interface

- Diagram of Air-Liquid-Interface exposure system and enlarged diagram of individual exposure cell



Diagrams Courtesy of Vitrocell Systems GmbH

Air-Liquid-Interface - Successes

- Measurement of Particle Deposition
 - Know the cell surface dose that results in the observed in-vitro/ex-vivo response?
 - For a range of solid particles 90 & 196 nm diameter solid particles – TEM using a single cell (Tippe et al., 2002)
 - 7 non-spherical solid particles 29 - 1600 nm diameter particles and 0.1 & 2 μm spherical solid particles - TEM (Comouth et al., 2013).
 - 8 spherical solid particles from 30 – 1005 nm in diameter – TEM in Vitrocell® six cell unit (Fujitani et al., 2015).
 - 5 solid particles from 0.5 – 1.6 μm in diameter – fluorescence - Vitrocell® 24/48 system (Steiner et al., 2017a,b).
 - 42nm & 40 μm – gravimetrically (Aufderheide et al., 2017) in a Cultex® radial flow unit.
 - mainstream tobacco smoke - spectrofluorometric quantification in a custom exposure system (Adamson et al., 2011)

Air-Liquid-Interface - Successes

- Measurement of Particle Deposition
 - Know the cell surface dose that results in the observed in-vitro/ex-vivo response?
 - Quartz crystal microbalances (QCM) - Successes
 - Changes in vibration frequency are correlated to amount of mass on the crystal (Sauerbrey, 1959) with 3 assumptions:
 - Mass deposited is small compared to the mass of the quartz crystal
 - Mass is rigid
 - Mass is evenly distributed over the crystal surface
 - Excellent sensitivity, depends on crystal thickness $\sim 10 \text{ ng/cm}^2$
 - Mainstream tobacco smoke (Adamson et al., 2012, 2013 a,b; Thorne et al., 2013; Majeed et al., 2014;)

Air-Liquid-Interface - Successes

- Measurement of Particle Deposition
 - Know the intended exposure concentration that results in the in-vitro/ex-vivo observed response?
 - Photometers
 - Light scattering is converted to aerosol mass or particle size distribution
 - Refractive index of aerosol will affect readings
 - TPM from mainstream tobacco smoke (Ritter et al., 2003)

- Know the cell exposure concentration that results in the in-vitro/ex-vivo observed response?
 - Chemical Identification
 - Vitrocell[®] 24/48 - mainstream tobacco smoke (Majeed et al., 2014)
 - Nicotine and metabolites from PBS below cells
 - Selected carbonyls from line exhaust
 - Cultex RFS module – mainstream tobacco smoke (Ishikawa et al., 2016)
 - Solanesol - modified insert with trapping solution at cell level
 - Acetaldehyde – modified insert with trapping solution at cell level



Air-Liquid-Interface - Successes

- Know the cell exposure concentration that results in the in-vitro/ex-vivo observed response?
 - Chemical Identification
 - 6 cell exposure module (manufacturer not specified) – mainstream tobacco smoke (Fields et al., 2017)
 - TPM – trapping solution at cell level
 - CO – online monitor



Air-Liquid-Interface - Challenges

- Liquid droplet aerosols (e-vapor product aerosols)
 - QCM not suited for measurement
 - Photometers - depending on exposure flowrate, aerosol concentration exceeds capacity
 - Significant amount of semi-volatile constituents (Ingebrethsen et al., 2012)
 - Differences in constituent refractive and constituent volatility could affect readings
- Relative Humidity > 75-80% RH (Zavala et al., 2017)
 - Most publications don't indicate RH
 - If they do, the RH existing the humidification system not exiting the exposure system is usually stated

Air-Liquid-Interface - Challenges

- Temperature = 37°C (Zavala et al., 2017)
 - Most publications indicate that cells are kept at 37°C but don't indicate the temperature of the exposure atmosphere
 - Difficult to raise temperature of exposure air due to small residence time in exposure system
- Thermophoretic Effect
 - Temperature difference between exposure air and cells creates a thermal barrier to deposition (Neilson et al., 2015; Higuchi et al., 2016)
 - The lower the exposure flowrates and the greater temperature difference the larger the thermal barrier

Air-Liquid-Interface - Challenges

- Equipment Operation

- Preliminary studies yield unexpected results in a Vitrocell 24/48 system for deposition of 3.2 μ m monodisperse fluorescent particles
 - Between 8,000 – 10,000 particles found on filters at the end of each row
 - Media/phosphate buffered saline below inserts was not used

Transwell ID	Particle Counts						
A1	132	A2	2	A3	166	A4	4
B1	162	B2	0	B3	144	B4	0
C1	170	C2	0	C3	139	C4	54
D1	53	D2	90	D3	1	D4	151
E1	1	E2	102	E3	0	E4	149
F1	4	F2	113	F3	0	F4	149



Air-Liquid-Interface - Challenges

- Equipment Operation
 - Lagrangian approach using Fluent[®] CFD software for air velocity profiles – added fluid channels below transwells to simulation



VC24_Channel_D2_2cc.mp4



Air-Liquid-Interface - Opportunities

- Quartz crystal microbalance with dissipation (QCM-D) - Challenges
 - Provides novel insight into viscoelastic properties of deposited mass (liquids and vapor: Vashist & Vashist, 2011)
 - With specific coatings has been used to measure specific volatile organic chemicals, ammonia, benzene, etc. (Vashist & Vashist, 2011)
- New Photometers
 - Dual lasers with polarized light designed for e-vapor products (Wang et al., 2017)
- Thermophoretic Effect
 - Use thermophoresis to drive deposition (heat the exposure air)
 - How realistic and how will cells respond?

Air-Liquid-Interface - Opportunities

- Electrostatic precipitation
 - Used for decades to collect ambient airborne particulate matter
 - For Bipolar charged 50 – 600nm particulate, 15-35% enhancement of particle deposition is possible (Savi et al. 2008)
 - Gaschen et al., 2010 and de Bruijne et al., 2009 have reported on custom made systems
 - Up to 90% deposition for 19-882nm particles (de Bruijne et al., 2009)
 - No cellular effects noted due to the electric field (de Bruijne et al., 2009)

Literature Cited

Adamson, J., Azzopardi, D., Errington, G., Dickens, C., McAughey, J., Gaça, M.D. Assessment of an in vitro whole cigarette smoke exposure system: The Borgwaldt RM20S 8-syringe smoking machine, *Chemistry Central Journal* 26;5:50, 2011, doi: 10.1186/1752-153X-5-50.

Adamson, J., Hughes, S., Azzopardi, D., McAughey, J., Gaça, M.D. Real-time assessment of cigarette smoke particle deposition in vitro, *Chemistry Central Journal*, 6:98, 2012.

Adamson, J., Thorne, D., Dalrymple, A., Dillon, D., Meredith, C. Cigarette smoke deposition in a Vitrocell® exposure module: real-time quantification in vitro using quartz crystal microbalances, *Chemistry Central Journal*, 7:50, 2012a

Adamson, J., Thorne, D., McAughey, J., Dillon, D. Meredith, C. Quantification of cigarette smoke particle deposition in vitro using a triplicate quartz crystal microbalance exposure chamber, *BioMed Research International*, (Article ID 685074), 2013b

Aufderheide, M., Heller, W-D., Krischenowski, O., Mohle, N., Hochrainer, D. Improvement of the Cultex® exposure technology by radial distribution of the test aerosol, *Experimental and Toxicologic Pathology*, 69:359-365, 2017.

Comouth, A., Saathoff, H., Naumann, K-H., Muelhopt, S., Paur, H-R., Leisner, T. Modelling and measurement of particle deposition for cell exposure at the air-liquid interface, *Journal of Aerosol Science*, 63:103-114, 2013.

Davies, C.N. Definitive equations for the fluid resistance of spheres. *The Proceedings of the Physical Society*, 57, part 4, No. 322, July 1, 1945.

Davies, C.N. (ed.), *Aerosol Science*. Academic Press, London, 1966.

De Bruijne, K., Ebeersviller, S., Sexton, K.G., Lake, S., Leith, D., Goodman, R., Jetters, J., Walters, G.W., Doyle-Eisele, M., Woodside, R., Jeffries, H.E., Jaspers, I. Design and testing of electrostatic aerosol in vitro exposure system (IEAVES): and alternative exposure system for particles, *Inhalation Toxicology*, 21:91-101, 2009.

Literature Cited

Fields, W., Maione, A., Keyser, B., Bombick, B. Characterization and Application of the VITROCELL VC1 Smoke Exposure System and 3D EpiAirway Models for Toxicological and e-Cigarette Evaluations, *Applied in Vitro Toxicology*, 3:2017, DOI: 10.1089/aivt.2016.0035.

Fujitani, Y., Sugaya, Y., Hashiguchi, M., Furuyama, A., Hirano, S., Takami, A. Particle deposition efficiency at air-liquid interface of a cell exposure chamber, *Journal of Aerosol Science*, 81:90-92, 2015.

Gaschen, A., Lang, D., Kalberer, M., Savi, M., Geiser, T., Gazdhar, A., Lehr, c.M., Bur, M., Dommen, J., Baltensperger, U., Geiser, M., Cellular responses after exposure of lung cell cultures to secondary organic aerosol particles, *Environmental Science and Technology*, 44:1424-1430, 2010.

Green, H.L. and Lane, W.R, *Particulate clouds: Dusts, smokes and mists*. 2nd ed., E.&F.N. Spon Ltd., London 1964.

Higuchi, M., Ledbetter, D.S., Morgan, S.D., McCullogh, S.D., White, P.A., Delvin, R.B., Zavala, J. Critical evaluation of air-liquid interface cell exposure systems for in vitro assessment of atmospheric pollutants, Abstract 1429, *Toxicologist*, March, 2016.

Hinderliter, P.M., Minard, K.R., Orr, G., Chrisler, W.B., Thrall, B.D., Pounds, J.G., Teeguarden, J.G. ISDD: A computational model of particle sedimentation, diffusion and target cell dosimetry for in vitro toxicity studies, *Particle and Fibre Toxicology*, 7:36, 2011.

Ingebretsen, B.J., Cole, S.K., Alderman, S.L. Electronic cigarette aerosol particle size distribution measurements. *Inhalation Toxicology*, 24: 976–984, 2012

Ishikawa, S., Suzuki, T., Nagata, Y., Analysis of cigarette smoke deposition within an in vitro exposure system for simulating exposure in the human respiratory tract, *Beitrag zur Tabakforschung International/Contributions of Tobacco Research*, 27:20-29, 2016.

Literature Cited

Majeed, S., Frentzel, S., Wagner, S., Kuehn, D., Leroy, P., Guy, P.A., Knorr, A., Hoeng, J., Peitsch, M.C. Characterization of the Vitrocell® 24/48 in vitro aerosol exposure system using mainstream cigarette smoke, *Chemistry Central Journal* 8:62, 2014

Neilson, L., Mankus, C., Thorne, D., Jackson, G., DeBay, J., Meredith, C. Development of an in vitro cytotoxicity model for aerosol exposure using 3D reconstructed human airway tissue; application for assessment of e-cigarette aerosol, *Toxicology In Vitro*. 29:1952-62, 2015.

Ritter, D., Knebel, J.W., Aufderheide, M. Exposure of human lung cells to inhalable substances: A novel test strategy involving clear air exposure periods using whole diluted cigarette mainstream smoke, *Inhalation Toxicology*, 15:676-84, 2003.

Sauerbrey, G. Verwendung von Schwingquarzen zur wagung dünner schichten und zur mikrowagung, *Z. Phys.*, 155:206-222, 1959

Savi, M., Kalberer, M., Lang, D., Ryser, M., Fierz, M., Gaschen, A., Ricka, J., Geiser, M., A novel exposure system for the efficient and controlled deposition of aerosol particles onto cell cultures, *Environmental Science and Technology*, 42:5667-5674, 2008

Steiner, S., Majeed, S., Kratzer, G., Vuillaume, G. Hoeng, J., Frentzel, S. Characterization of the Vitrocell 24/48 aerosol exposure system for its use in exposures to liquid aerosols, *Toxicology in Vitro*, 42:263-272, 2017a.

Steiner, S., Majeed, S., Kratzer, G., Hoeng, J., Frentzel, S. A new fluorescent-based method for characterizing in vitro aerosol exposure systems, *Toxicology in Vitro*, 38:150-158, 2017b.

Literature Cited

Thorne, D., Kilford, J., Payne, R., Adamson, J., Scott, K., Dalrymple, A., Meredith, C., Dillon, D. Characterization of a Vitrocell® VC10 in vitro smoke exposure system using dose tools and biological analysis, *Chemistry Central Journal*, 7:146, 2013.

Thomas, D.G., Smith, J.N., Thrall, B.D., Baer, D.R., Jolley, H., Munusamy, P., Kodali, V., Demokritou, P., Cohen, J., Teeguarden, J.G. ISD3: a particokinetic model for predicting the combined effects of particle sedimentation, diffusion and dissolution on cellular dosimetry for in vitro systems, *Particle and Fiber Toxicology*, 15:6, 2018. doi: 10.1186/s12989-018-0243-7

Tippe, A., Heinzmann, U., Roth, C. Deposition of fine and ultrafine aerosol particles during exposure at the air/cell interface, *Aerosol Science*, 33:207-218, 2002.

Vashist, S.K. and Vashist P. Recent advances in quartz crystal microbalance-based sensors, *J. of Sensors*, doi:10.1155/2011/571405. Zavala, J., Greenan, R., Krantz, Q.T., DeMarini, D.M., Higuchi, M., Gilmour, M.I., White, P.A. Regulating temperature and relative humidity in air-liquid interface in vitro systems eliminates cytotoxicity resulting from control air exposures, *Toxicology Research*, 6:448-459, 2017.

Wang, Q.; Li, W.; Lipowicz, P.; Dunkhorst, W.; Koch, W., "A Light Scattering Sensor for the Analysis of Electronic Cigarette Aerosols". In: Verbal presentation at the 71st Tobacco Science Research Conference (TSRC), Bonita Springs, Florida, November 28 – December 1, 2017, 2017.

Zavala, J., Greenan, R., Krantz, Q.T., DeMarini, D.M., Higuchi, M., Gilmour, M.I., White, P.A. Regulating temperature and relative humidity in air-liquid interface in vitro systems eliminates cytotoxicity resulting from control air exposures, *Toxicological Research*, 6:448-459, 2017.

Thank you



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