How to get started, collect sterile milk samples, culture bacteria and diagnose results











Treatment Decisions for Clinical Mastitis:

Part One: Will Antibiotics Help the Cow?

Part Two: Using Cultures to Make Selective Treatment Decisions

How to Set Up Your On Farm Laboratory

Selecting Culture Media

Mastitis Severity Scoring

How to Collect an Aseptic Milk Sample

How to Set Up Culture Plates

How to Read Culture Plates

How to Identify Staphylococcus Species Using Selective Agars

How to Identify Streptococcus Species Using Selective Agars

How to Identify Gram-Negative Species

On Farm Culturing: What Can Go Wrong?

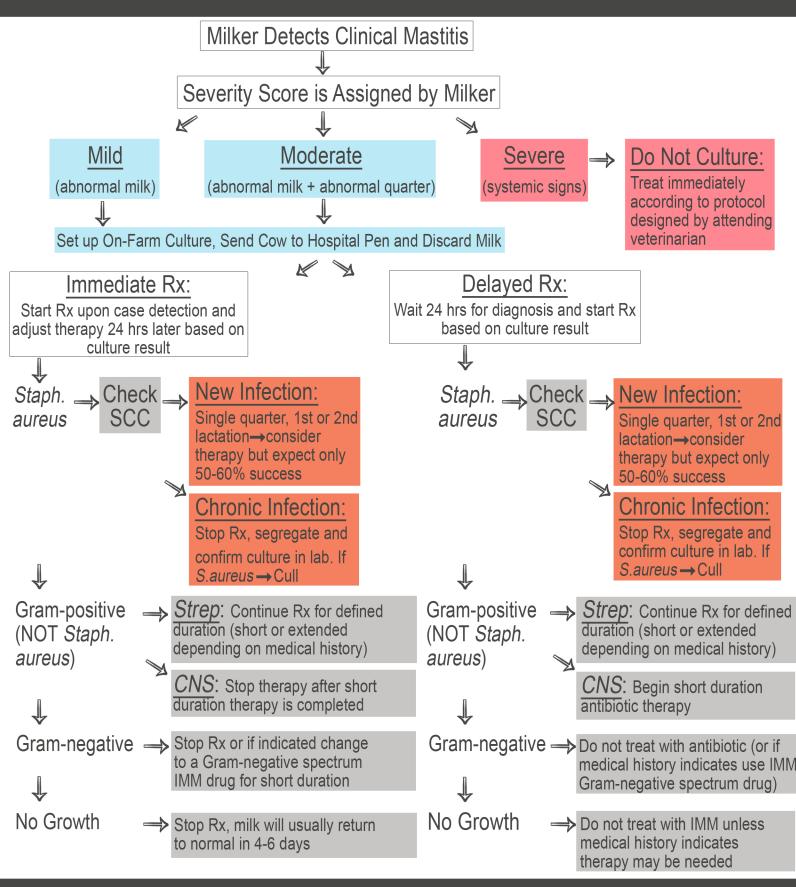






















On farm culture systems require a **designated workspace**. Ideally, this should be a clean, well-lit room in a low-traffic area. There should be sufficient counter space and storage that are easily disinfected. Food should not be allowed in this area for health reasons.

Incubator

- o A small **incubator** should be purchased to create an environment ideal for bacterial growth.
- o Keep the incubator at 37*C or 98.6*F (body temperature).
- o A thermometer should be kept inside the incubator at all times, and should be checked daily.
- o The humidity should be maintained at 75% by placing a dish of water inside the incubator. Water level should be checked daily and replenished as needed.

Refrigerator

o A **refrigerator** should be purchased for storing media plates and saved milk samples. This refrigerator should not be used for human or animal food.

Equipment

- o On-farm cultures use **sterile swabs** to plate milk samples instead of the sterile loops used in milk quality labs. The estimated plating volume of swabs is 0.1 mL if the swab is dipped in the milk sample for 10 seconds prior to plating.
- o Disposable gloves should be worn at all times when handling lab materials.
- o Media plates
- o Gauze squares soaked in 70% alcohol for disinfecting teats and the counter surface
- o Single-use milk sample vials
- o Racks for holding sample vials
- o Permanent marker
- o Biohazard bags
- o Bleach
- o A **cooler** with ice for transporting samples from the cow to the lab area.

Plate Disposal

- o One more consideration for lab set-up is waste disposal. Use orange biohazard bags whenever discarding infectious materials such as milk samples and culture plates.
- o This lab waste must be disposed according to your local regulations. In some locations, plates may be flooded with bleach and then disposed of normally
- o If you have questions about disposal protocol in your area, ask your herd veterinarian.



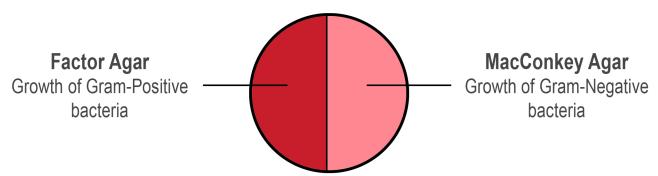




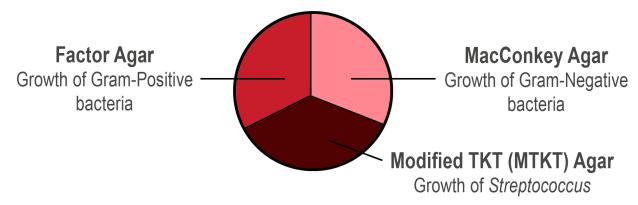




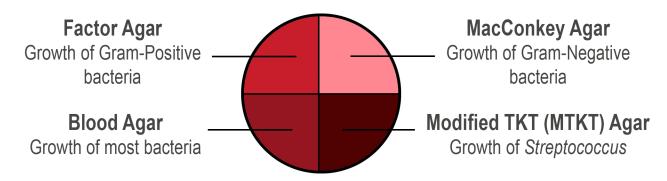
Biplate
Results: Gram-positive, Gram-negative bacteria, No growth or contaminated



Triplate
Results: Staphylococcus spp., Streptococcus spp., Gram-negative bacteria,
No growth or contaminated



Quad plate
Results: Staphylococcus spp., Streptococcus spp., Gram-negative bacteria,
No growth, contaminated or others.













treatment decisions. The degree of illness and the symptoms present will depend on many factors, such as the nutritional or immune status of Clinical mastitis is an udder infection that shows symptoms which are visible. The level of infection, or severity, can help herd managers make the cow, which pathogen is responsible for the inflammation, and a range of environmental factors such as cleanliness, humidity and ambient temperature. Moderate to severe clinical cases can be very painful and unpleasant for the cow.



Milk has a watery appearance, **ABNORMAL MILK** flakes or clots.



swelling, heat, hardness, red-ABNORMAL UDDER Signs of inflammation: ness or pain.



Reduction in milk, fever, lack **ABNORMAL BEHAVIOR** of appetite, sunken eyes, diarrhea, dehydration or reduction in mobility.



OW MILK QUALITY



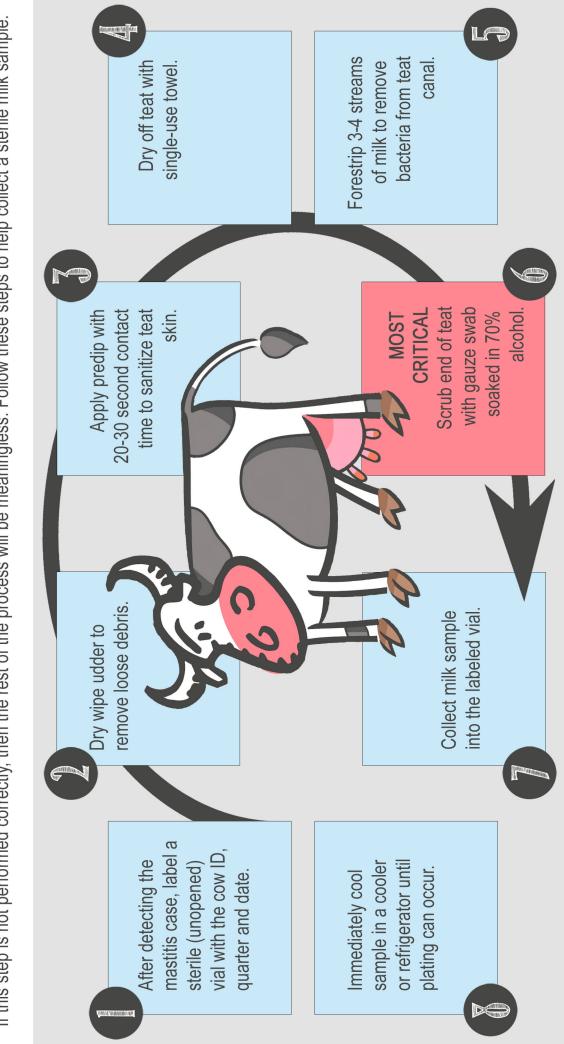








If this step is not performed correctly, then the rest of the process will be meaningless. Follow these steps to help collect a sterile milk sample. The entire process of performing on farm culturing is absolutely dependent on how well the milk sample was collected.











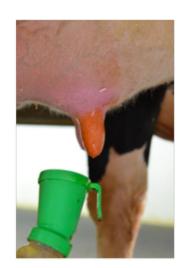


After detecting the mastitis case, label a sterile (unopened) vial with the cow ID, quarter and date.



- Dry off teat with single-use towel.
- Apply predip with 20-30 second contact time to sanitize teat skin.





- Dry off teat with single-use towel.
- Forestrip 3-4 streams of milk to remove bacteria from teat canal.







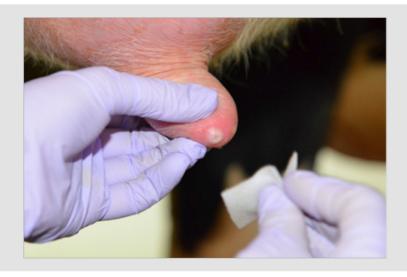






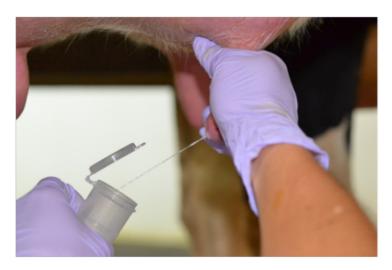


MOST CRITICAL Scrub end of teat with gauze swab soaked in 70% alcohol.





Collect milk sample into the labeled vial. Do not touch any inner part of the vial to avoid contamination of the sample.





Immediately cool sample in a cooler or refrigerator until plating can occur.













Before plating, check that all equipment is present and accessible:

- o Culture plates
- o Sterile swabs
- o Milk samples and sample rack
- o Clean disposable gloves
- o Gauze soaked in 70% alcohol

Clean work space with gauze and 70% alcohol to eliminate environmental bacteria that otherwise might contaminate the culture plates. When setting up culture plates be sure to always wash your hands and wear clean disposable gloves.

Thaw any frozen samples to room temperature. This will require setting the samples out on the lab bench for at least an hour prior to plating. Do not thaw frozen samples using hot water or the microwave.

Label the side of the culture plate with cow ID, affected quarter and the date.

Thoroughly mix the milk sample by inverting it several times. Dip a sterile cotton swab into the sample and wait 5 seconds for it to become saturated with milk. Spread the swab over the media surface using a back-and-forth motion from the top down. Redip the swab in the milk sample for each section of the culture plate. Immediately cover the plate and cap the milk sample after plating to avoid contamination.

Freeze milk samples so they are available for confirmatory testing later.

Lay the plates flat on the work surface for 5-10 minutes to allow the milk to soak into the agar.

Invert all plates so the agar side is up so that condensation that collects on the lid will not drip onto the agar and disrupt bacterial growth.

Place the upside-down plates in the incubator and incubate at 37 degrees Celsius (98 degrees Fahrenheit) for 24 hours. The bacteria will need a minimum of 24 hours to grow before reading results.

Throw away any garbage and disinfect the work space.











Have all materials ready to use:
Milk samples
Culture plates
Sterile swabs
Gloves
Gauze soaked in 70% alcohol
Biohazard bags



Label the side of a new on-farm culture plate with:
Cow ID
Affected quarter
Date



Thoroughly mix the milk sample by inverting it several times.













Dip a sterile swab in the milk sample and roll it until swab becomes completely saturated with milk.



Inoculate one section of a culture plate using a zigzag streaking pattern.



Re-dip the swab in the milk sample before inoculating each section of the culture plate.













Inoculate the resting sections of the culture plate.



Place the inoculated culture plates in the incubator upside down (lid facing the shelf) and incubate plates for 18 to 24 hours at 37°C.













The purpose of performing on farm culturing is not only to grow bacteria but to ensure that our diagnosis is correct so that we can determine if antibiotic treatment will be helpful.

First Step: Assessing Bacterial Growth

Bacterial growth takes a minimum of 24 hours in incubation before the plate can be observed. After 24 hours, the first question to ask is: **Is there any growth on the plates?**

No Growth

There are several reasons this happens. First, the quarter may not be infected or the infection may already have been cleared by the body's immune system. Also, improper handling or lab errors may cause no growth. Lastly, fastidious organisms that require special media, such as Mycoplasma, may cause infection but will not grow on standard culture plates.

Second Step: Interpretation of Bacterial Growth

If there is growth on the plate, the second question to ask is: **How many different types of bacterial colonies are there?**

Contaminated

If there are more than 2 types of colonies, the plate has been contaminated. When this happens, the quarter may be resampled using proper collection technique and cultured again.

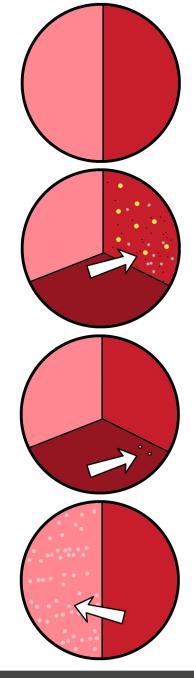
If there are 1 to 2 colony types, the number of 'colony forming units' should be evaluated. A 'cfu' is a small circular, often raised growth of bacteria.

Non-significant Growth

Fewer than 3-5 cfu's per colony type signifies non-significant growth. This means that the bacteria present on the culture plate are too few in number to be the cause of mastitis.

True Infection

When there are 3-5 or more cfu's per colony type, there is a true infection, and identification can be performed. Sometimes there may be a true infection that has a contaminant. As long as the contaminant has non-significant growth, you can still identify the true infection.









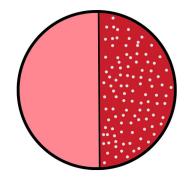




Determining the cause of mastitis is important because not all cases of mastitis benefit from antibiotic therapy. For instance, Gram-Positive bacteria, such as Staphylococcus aureus and coagulase negative behave differently in the cow and have different responses to therapy. Being able to identify between species can help us make appropriate treatment decisions for managing mastitis in our herds. One way to do that is through on farm culturing.

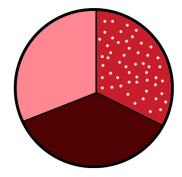
Biplate

Gram-Positive bacteria (growth only on Factor or Blood agar)



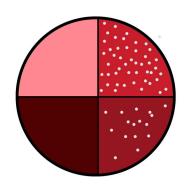
Triplate

Staphylococci (growth only on Factor or Blood agar)

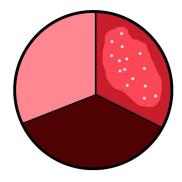


Quadplate

Staphylococci (growth on Blood Agar and Factor agar)

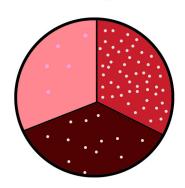






Contamination

Contaminated plate (growth on all agars)









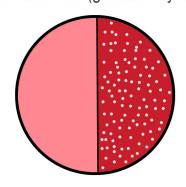


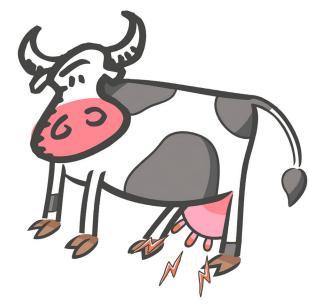


Streptococcus species are a common cause of mastitis and frequently associated with high somatic cell counts, and in some cases clinical mastitis. Streptococci are gram positive organisms that also grow in the environment. Learn how to identify Streptococci on a biplate, triplate and quadplate using selective agars in your on-farm culturing lab.

Biplate

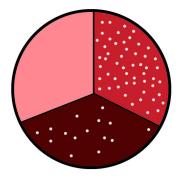
Gram-Positive bacteria (growth only on Factor or Blood agar)





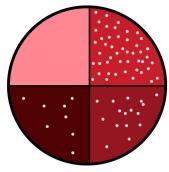
Triplate

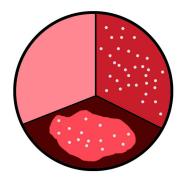
Streptococci (growth on both Factor and MTKT agars) Streptococcus agalactiae (zone of hemolysis in MTKT)



Quadplate

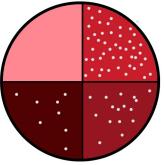
Streptococci (growth on Blood Agar, Factor and MTKT agar)

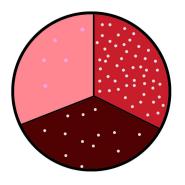




Contamination

Contaminated plate (growth on all agars)











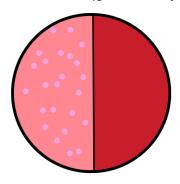




Gram negative organisms cannot be differentiated at the genus level (such as E. coli, Klebsiella or Enterobacter) on the agar plates used in on-farm cultures. However, they can be identified as lactose negative or lactose positive by what color they ferment lactose in MacConkey agar. Gram negative infections often resolve on their own. Therefore, it is not always necessary to treat with antibiotics. Remember, it is always advisable to consult your local veterinarian when making these decisions.

Biplate

Gram-Negative bacteria (growth only on MacConkey and blood agars)

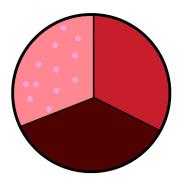


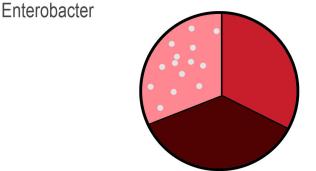


Triplate

Lactose-postive (pink growth on MacConkey agar) Lactose-negative (white/yellow growth on MacConkey agar)

E. coli Klebsiella

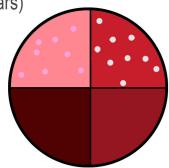




Quadplate

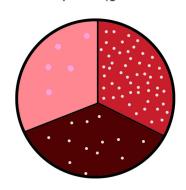
Gram-Negative bacteria (growth only on MacConkey

and blood agars)



Contamination

Contaminated plate (growth on all agars)













Contamination

One problem that can easily occur for on-farm culture labs is contamination during sample collection, handling or plating. Contamination can be difficult to detect on selective media, since many

contaminants are not able of grow. Prevention is key in reducing the number of plates that become contaminated.

Contamination during sample collection may occur if udders are not properly disinfected prior to sampling, if the teat, the cow's tail, or another source of manure contacts the sample vial, or if the vial is not closed promptly after sampling.

Contamination during <u>sample handling</u> can occur if the sample is not placed in a cooler during transport to the lab, or if it is allowed to sit out for greater than one hour prior to plating.

Contamination during <u>plating</u> can happen if staff do not wear clean disposable gloves, if sterile swabs are left uncovered or contact non-sterile material, or if plates are not covered immediately after plating.

Benchmark: If more than 5% of the culture plates are contaminated, procedures should be evaluated for aseptic technique, and the appropriate changes or training should be performed.

Failure of Quality Control

Another problem that can influence the value of culture results is failure of quality control lab processes.

Incubator <u>temperature</u> should be maintained at 37 degrees Celsius or 98.6 degrees Fahrenheit. If the incubator temperature is too low or too high, disease-causing bacteria will not grow as well. This can lead to no-growth samples or mis-diagnoses.

Incubator <u>humidity</u> must remain high as well. A container of water should be kept in the incubator and refilled often to maintain moisture levels conducive to bacterial growth.

Benchmark: On-farm culture results should be compared to milk quality lab results at specified intervals to evaluate the quality of on-farm interpretation. They will not be identical but they should agree in general about 80% of the time.









Contamination

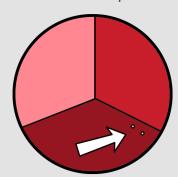


Over-interpretation of Bacterial Growth

In real diagnostic labs about 25-40% of milk samples from cows with clinical mastitis result in no-growth or non-significant growth. It is common for a few bacteria to get picked up from teat skin or during sampling, but bacteria on contaminated plates and non-significant growth are not considered the cause of mastitis. Antibiotics should not be given based on contaminated growth or non-significant growth.

Benchmark: Only when you have 1 to 2 types of colonies with at least 3-5 colony-forming-units can you be confident in your bacterial identification and choice of treatment.

Non-significant Growth
Fewer than 3-5 cfu's per colony type



Failure to Use Information Properly

The most common problem at on-farm culture labs is that farmers do not use the culture results for making treatment decisions. The value of any diagnostic test is based on the economic value of the intervention that one makes. If the culture results are not taken into account during real-time decision making, the lab is not having the impact it should on the farm, antibiotic use and costs will not be reduced, and, therefore, the value of culturing is lost.

Benchmark: Work with your veterinarian to design and implement appropriate treatment protocols for your farm. Remember, your local veterinarian and milk quality lab are here to help you. Be sure to contact them for support and accountability in maintaining the quality of your on-farm culture system.









